

Regulation of Photosynthesis during the Light Period in CAM Plants —Evaluation by a Gas-Phase O₂ Electrode and a Compensating Infrared CO₂ Analysis System—

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The CO₂ dependent O₂ evolution during the light period, at which the exogenous CO₂ uptake was suspended, in 12 CAM plants (including pineapple (*Ananas comosus*)) was evaluated with a gas-phase oxygen electrode. At 5% CO₂, the rate of photosynthetic O₂ evolution in pineapple was saturated at 1 500 μmol m⁻² s⁻¹ PPFD and the maximum rate was 60 μmol m⁻² s⁻¹, which was 10 times those obtained at ambient CO₂ conditions with the CO₂ exchange system and significantly higher than the other CAM plants. At the saturated PPFD, the O₂ evolution in pineapple substantially increased with increasing CO₂ concentration up to 3% and decreased above 4%. However in the other CAM plants, such increment was small. By the use of a novel compensating CO₂ gas exchange system, the rate of CO₂ exchange in pineapple at very high CO₂ was measured and found the enhancement of CO₂ uptake in both light (Phases III and IV) and dark (Phase I) periods. Based on the results obtained, possibilities of the further increases in CO₂ uptake in CAM plants, especially of pineapple, are discussed in terms of stomatal functions and malate storage capacities.

Keywords: CO₂ dependent O₂ evolution, crassulacean acid metabolism, high CO₂, pineapple, stomatal closure

INTRODUCTION

In pineapple cultivation, at least 2 years is needed from transplanting to harvest under natural conditions. The use of plant hormones such as indoleacetic acid and ethylene accelerates the floral differentiation and can reduce the cultivation period at most 6 months. In general, the growth rate of pineapple is extremely slow due to its crassulacean acid metabolism (CAM), as compared with C₃ and C₄ plants (Nose et al., 1986). Recently, some farmers in Okinawa, Japan, are cultivating pineapple in greenhouses to promote growth rate and simultaneously to improve fruit quality. Although they can improve fruit quality in winter and early spring, they still unsuccessful to shorten the cultivation period. Main reason of this is that most farmers do not understand pineapple as a CAM plant and its tremendous performances in physiological and biochemical aspects are not accounted for the actual

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cultivation. For example, some farmers close the greenhouses in order to raise night temperature, irrespective of the evidence that pineapple plants absorb CO₂ at night through CAM. This makes the pineapple leaf into CO₂ starved state at the end of night (Kawamitsu and Nakayama, 1997).

According to Osmond (1978), the gas exchange and stomatal behaviour of CAM plants were divided into four phases, i.e. Phase I with CO₂ uptake in darkness, Phase II with CO₂ uptake in the early morning, Phase III with stomatal closure during the middle of day and Phase IV with CO₂ uptake again in the later afternoon. During Phase III, when stomata close tightly, malic acid previously formed in the dark is decarboxylated and refixation of CO₂ by ribulose 1,5-bisphosphate carboxylase/oxygenase (Rubisco) takes place. As a result of stomatal closure, the conventional gas exchange apparatus with an infrared gas analyzer can not be used to evaluate the CO₂ fixation by Rubisco in CAM leaf. Fortunately, a gas-phase O₂ electrode system can detect the O₂ evolution from CAM leaves (Thomas and Andre, 1987; Maxwell et al., 1998). Even if the stomata are closed, CO₂ with very high concentrations (1–10%) can penetrate through waxy epidermis or cutting edge, so that CO₂ dependent O₂ evolution operating in the mesophyll cells may be detected with a gas-phase leaf disc O₂ electrode (Delieu and Walker, 1981).

Although the rise in CO₂ is a world wide phenomenon, few studies on the effects of CO₂ enrichment for pineapple plants were undertaken (Zhu et al., 1997, 1999). Elevated CO₂ would be expected to have little effect on dark CO₂ fixation because it is mediated by phosphoenolpyruvate carboxylase (PEPCase), which is assumed to be CO₂ saturated at near-ambient CO₂ levels (Ting, 1994). However, Zhu et al. (1997, 1999) demonstrated that PEPCase in pineapple leaf is not CO₂-saturated at ambient CO₂ levels and 25°C night temperature.

The goal of this research was to establish an ideal technique for the cultivation of pineapple with reducing the growth period at least to 12–13 months. We expect that an increase in leaf CO₂ exchange rate may be an important factor to increase the carbon accumulation in pineapple, a CAM plant. In this paper, we examined the responses of O₂ evolution to photosynthetic photon flux density (PPFD) and CO₂ concentration during light period in which exogenous CO₂ uptake was suspended (Kluge and Ting, 1978). In addition, to ensure the effects of very high CO₂ concentration on the CO₂ uptake during light period, we conducted the measurements of CO₂ exchange by a new, compensating gas-exchange system with an infrared gas analyzer (Graan and Boyer, 1990; Kawamitsu and Boyer, 1999).

MATERIALS AND METHODS

Plant materials. We examined 12 CAM species: *Ananas comosus* (L.) Merr. Smooth cayenne c.v. N67-10 (pineapple), *Crassula argentea*, *Dendrobium ekapol* cv. Panda No. 1, *Hoya carnosa* R. Br., *Kalanchoe blossfeldiana* Poelln., *K. daigremontiana* R. Hamet & Perrier, *K. fedtschenkoi* Hamet & Perr., *K. gastonis-bonniere* Hamet et Perr., *K. pinnata* Pers., *Peperomia incana* A. Dietr., *Sedum praealtum*, *Vanilla fragrans* Ames. These plants were propagated by adventitious plantlets obtained from a local botanical garden in Okinawa, Japan. Plants were grown in 1/5 000 a pots containing vermiculite. Pineapple plants were propagated by crowns obtained from Okinawa Experiment Station, Nago City, Okinawa, Japan. They were grown in 1/5 000 a pots containing red soil (Kunigami Maji, pH 5.8). Five hundred mL of modified Hoagland's solution was supplied twice a week. Composition of the solution was 6 mM Ca(NO₃)₂•4H₂O, 6 mM KNO₃, 2 mM KH₂PO₄, 2 mM MgSO₄•7H₂O, 25 μM H₃BO₃, 10 μM MnSO₄•4H₂O, 2 μM ZnSO₄•7H₂O, 0.5 μM CuSO₄•5H₂O, 0.5 μM H₂MoO₄, and 0.1 mM FeC₆H₅O₇. Plants were grown in a glasshouse under natural light

regimes. During the experiments, all plant materials were transferred into a growth chamber in photosynthesis measurement room, setting at $500 \mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD and $30/25^\circ\text{C}$ day/night temperature.

Measurements of CO_2 dependent O_2 evolution. The CO_2 dependent O_2 evolution was measured by a leaf-disc oxygen electrode system (Delieu and Walker, 1981, 1983; Buah et al., 1999; Kawamitsu and Boyer, 1999; Kawamitsu et al., 2000). According to preliminary experiments, the maximum photosynthetic rates were obtained at the uppermost, fully expanded leaves. Approximately 8 cm^2 leaf discs from leaves were cut and their fresh weights were determined. Then the leaf disc was placed in the electrode chamber and the volume of the chamber was calibrated after Delieu and Walker (1981, 1983). The CO_2 in the chamber is rapidly exhausted by photosynthesizing disc unless it is replenished (Delieu and Walker, 1981, 1983). Generally, a carbonate/bicarbonate buffer was recommended as a source of CO_2 in the system. However, we used gas mixture because CO_2 level could be altered rapidly without opening the chamber (Kawamitsu and Boyer, 1999; Kawamitsu et al., 2000). The procedure employed was as follows; the tissue was illuminated in the presence of gas mixture for at least 5 min prior to sealing the chamber and measuring O_2 exchange. After sealing the chamber, to get the trace of O_2 exchange on a chart of the recorder it ran for approximately 50 s. Subsequently, the chamber was flushed with the new gas mixture for 5 min and then the O_2 evolution was re-measured in the same manner. If the O_2 evolution at 10 min was the same as that at 5 min, the rate was considered stable and the conditions could be changed for the next set of measurement. The air humidity of chamber was maintained at near saturation to avoid desiccation of the leaf disc. Photosynthetic O_2 evolution as a function of CO_2 concentration and irradiance was determined at 21% O_2 . The Björkman lamp was used as a light source. The irradiance levels were altered by the use of neutral density glass filters, which were exchanged in the light source housing (LS-2, Hansatech). Irradiance was reduced stepwise from high PPFD ($1500 \mu\text{mol m}^{-2} \text{s}^{-1}$) to darkness in nine steps. Total darkness was obtained by wrapping the electrode chamber in two layers of aluminum foil. Equilibration time to establish a steady rate of O_2 evolution under a new PPFD in the presence of CO_2 required approximately 5 min for the CAM plants used. Because disc temperature rose by illumination, the water temperature circulating the electrode was maintained at approximately 23.5°C , so that the tissue temperature in the chamber was at 25°C after sealing. This temperature was employed for all the O_2 electrode experiments reported here. In the case of the dark respiration, the temperature of water bath was adjusted at 25°C since the temperature of the water bath and the electrode chamber was the same. For the measurement of CO_2 dependent O_2 evolution, CO_2 concentration of the incoming air was controlled by mixing CO_2 -free air (containing 21% oxygen and balanced nitrogen) with 10% CO_2 (containing 21% oxygen and balanced nitrogen). The CO_2 concentration was increased stepwise from 0 to 5% at 25°C disc temperature and $1500 \mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD. Each measurement was repeated 3 times and the higher value was plotted.

Measurements of CO_2 exchange. Diurnal changes in CO_2 exchange rates at normal air were determined with an infrared gas analyzer (Model Li-6251, Li-Cor) in an open system (Du et al., 1996; Kawamitsu et al., 1999a, b). After the plant was set in the assimilation chamber, gas exchange rate was monitored for about 1 week. Thereafter, the 24 h continued data sets were taken in the stage in which gas exchange rate was stabilized. The CO_2 exchange rates at very high CO_2 were measured at University of Delaware, College of Marine Studies, Lewes, Delaware 19958, U.S.A. with a semi-closed compensating system described in elsewhere (Graan and Boyer, 1990; Kawamitsu and Boyer, 1999). This system enables to increase the ambient CO_2 concentration up to 7% (70 000 ppm). The leaf area used for the experiments

was 120 cm². Gas exchange over 24 h was repeated twice at each CO₂ level for different plant with comparable results.

Organic acid and chlorophyll content. Organic acids were determined on the same leaf used in the measurement of CO₂ exchange under ambient CO₂ conditions. Leaf discs (2 cm²) were weighed and then boiled for 10 min, thereafter ground in 10 mL glass-distilled water with a Potter-type tissue homogenizer. The mixture was centrifuged at 12 000 × *g* for 10 min, and then filtered with the membrane filter (0.4 μm in diameter). Organic acid contents were determined with the liquid chromatography method (Model LC-6A, CDD-6A ; Shimadzu). Other leaf discs used for CO₂ dependent O₂ evolution were weighed and then ground with sea sand in a chilled mortar and pestle. Chlorophyll contents were determined with a spectrophotometer after extracting with 80% acetone (Arnon, 1949).

RESULTS AND DISCUSSION

Light response curves of CO₂ dependent O₂ evolution in pineapple and other CAM plants are shown in Fig. 1. All measurements were conducted during Phase III of CAM cycle (Osmond, 1978). Intercellular CO₂ concentration in CAM plants was often observed at 1–2% during Phase III (Cockburn, et al., 1979 ; Spalding et al., 1979). Thus we measured photosynthetic O₂ evolution at 5% CO₂, which was higher than intercellular CO₂ concentra-

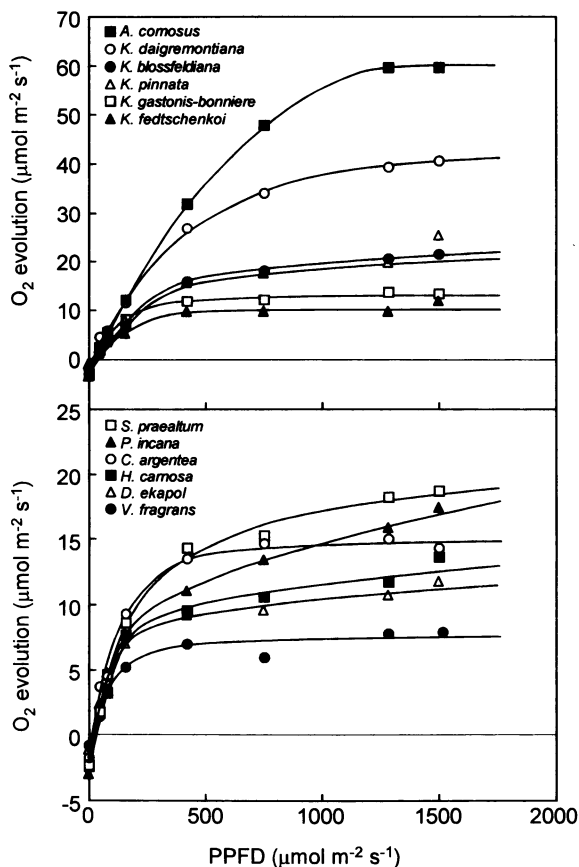


Fig. 1 Light response curves of O₂ evolution in CAM plants. Measurements were made at 5% CO₂.

tions reported. The rates of photosynthetic O₂ evolution in other CAM plants were also measured under the same conditions. Pineapple showed the highest O₂ evolution rate at higher PPFD (Fig. 1). The light saturation point of the O₂ evolution was approximately 1 500 μmol m⁻² s⁻¹ PPFD. *K. daigremontiana* showed an intermediate rate of O₂ evolution between pineapple and the other CAM plants (below 20 μmol m⁻² s⁻¹). In addition, the light saturation points were lower than 500 μmol m⁻² s⁻¹ PPFD in those CAM plants. Sixty μmol m⁻² s⁻¹ of O₂ evolution in pineapple was 7.5–12 times of the value of 5–8 μmol m⁻² s⁻¹ obtained at ambient CO₂ conditions with the conventional CO₂ exchange system (Fig. 6).

The photosynthetic O₂ evolutions were also expressed on a chlorophyll content basis (Fig. 2). Pineapple still showed higher photosynthetic rate than the other CAM plants. However, *K. gastonisbonniere*, which had a low value on a leaf area base in Fig. 1, showed the highest rate on a unit of chlorophyll base (Fig. 2). Growth rate of this species was higher than those of the other CAM plants, except pineapple (data not shown).

During the decarboxylation phase of CAM mode (Phase III), the release of CO₂ from stored malic acid generates high intercellular CO₂ concentration in the stomata-closed leaf, so that photosynthetic O₂ evolution occurs even when measured at CO₂ free, ambient air (Fig. 3). Although some CO₂ had escaped when the disc sample was punched from the intact leaf, O₂ evolution at 1 500 μmol m⁻² s⁻¹ PPFD was approximately 10 μmol m⁻² s⁻¹, which was one-sixth of the maximum O₂ evolution at 5% CO₂ (Fig. 1). When CO₂ concentration around the leaf disc was increased to the normal ambient air (0.03%), the rate of photosynthetic O₂ evolution remained unchanged (Fig. 3). It meant that the stomatal conductance was unusually low during Phase III, so that 0.03% CO₂ could not penetrate through epidermis or alternatively, disc intercellular CO₂ concentration was higher than the CO₂ concentration in the electrode chamber. Usually, in C₃ and C₄ plants, photosynthetic rate increases with increasing ambient CO₂ concentration. But the saturation point in C₄ species is lower than that in C₃ species because of CO₂ concentrating mechanism based on PEPCase and the lack of photorespiration (Long, 1999). In CAM plants, if the intercellular CO₂ concentration were maintained at 1–2% during Phase III, photosynthetic rate may not be influenced with altering CO₂ concentration up to 2% around the leaf disc. In pineapple, however, photosynthetic O₂ evolution was increased with increasing CO₂ up to 3% and subsequently decreased above 4% CO₂ (Fig. 4). Cockburn et al. (1979) showed 0.5% of intercellular CO₂ concentration in pineapple when leaves were illuminated. As shown in the present study (Fig. 4) and

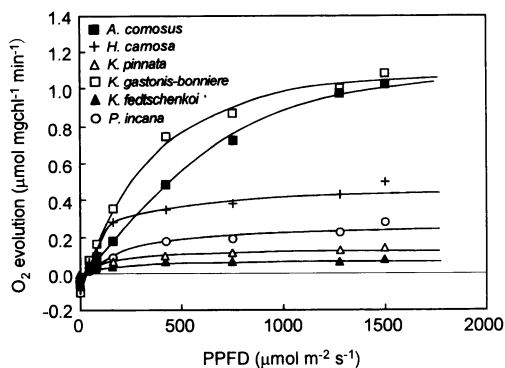


Fig. 2 Light response curves of O₂ evolution in CAM plants.

Measurements were made at 5% CO₂ and expressed on a chlorophyll base.

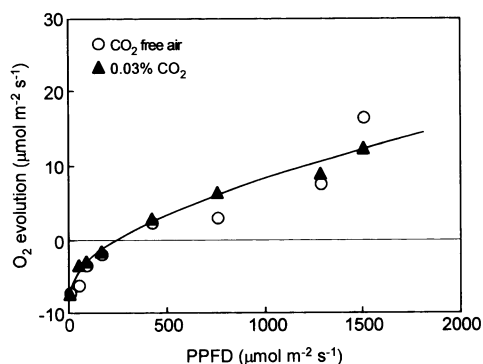


Fig. 3 Light response curves of O₂ evolution in pineapple at CO₂ free air and 0.03% CO₂.

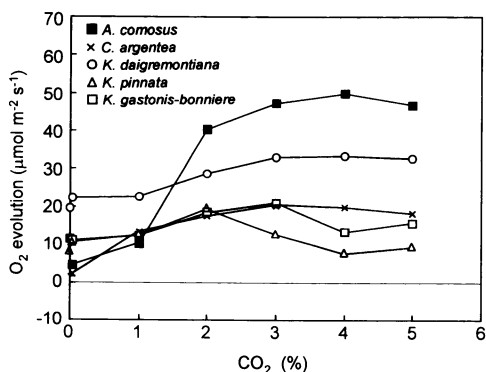


Fig. 4 Effects of CO₂ concentration on photosynthetic O₂ evolution in CAM plants. Measurements were made at 1 500 μmol m⁻² s⁻¹ PPFD, 21% O₂ and 25°C disc temperature.

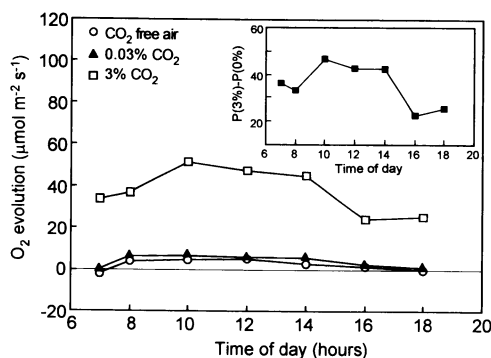


Fig. 5 Time courses of photosynthetic O₂ evolution at different CO₂ concentration in pineapple. The inset indicates the difference in O₂ evolution between at 3% CO₂ and CO₂ free air.

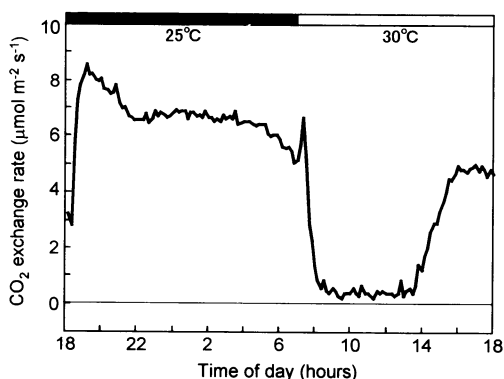


Fig. 6 Diurnal courses of CO₂ exchange rate at ambient CO₂ concentration in pineapple. Day/night temperature was set at 30/25 °C, and PPFD was 600 μmol m⁻² s⁻¹ during the light period.

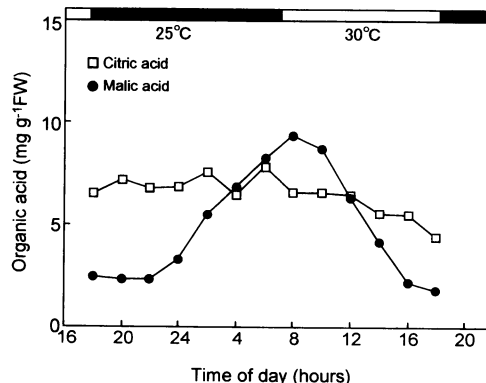


Fig. 7 Diurnal courses of organic acid contents in pineapple leaf. Conditions were the same as in Fig. 6.

some other studies (Cockburn et al., 1979; Spaldings et al., 1979), we assume that the intercellular CO₂ concentration in pineapple in Phase III is at around 0.5%. Moreover, maximum O₂ evolution at high CO₂ concentration in pineapple is higher than those of other CAM plants. Due to insufficient CO₂ diffusion at the most inner side of thick leaf in pineapple, only the mesophyll cells near the stomatal cavities may be actively fix CO₂ from the ambient air. In the other CAM plants, the degrees of increment in photosynthetic O₂ evolution with increasing CO₂ concentration were small, indicating that photosynthesis are already saturated at the intercellular CO₂ concentration. If so, the intercellular CO₂ concentration in those CAM plants was probably lower than that of pineapple.

Diurnal changes in photosynthetic O₂ evolution at very high CO₂ concentration were measured during the light period (Phases II, III and IV) (Fig. 5). As a comparison, the diurnal course in CO₂ exchange of the different leaf of the same plant at ambient CO₂

conditions was also measured (Fig. 6). In pineapple, CO₂ uptake occurred during Phases I, II and IV, and its mode was classified as full CAM type (Kluge and Ting, 1978). Maximum CO₂ exchange rate in Phase I was approximately 7 μmol m⁻² s⁻¹ (Fig. 6) and a little CO₂ uptake was detected during Phase III. Diurnal changes in malic and citric acids of the same leaf exhibited a typical CAM pattern (Fig. 7). At the end of Phase I, malate content reached the peak value, and thereafter decreased to the minimum value at Phase IV. However, Fig. 5 showed that O₂ evolution occurred throughout the light period (Phases II, III and IV) and furthermore, photosynthetic O₂ evolution at 3% CO₂ was 8–10 times higher as compared with those at 0.03% CO₂. There were no differences in photosynthetic O₂ evolution between at 0.03% CO₂ and CO₂ free air because of the difficulties of CO₂ penetration through the epidermis of pineapple leaf. At 3% CO₂, the photosynthetic O₂ evolution had a peak at around 10:00 (Phase III) and decreased at Phases II and IV. Under CO₂ free, ambient air, there are no carbon sources around the leaf disc, but O₂ evolution occurred during the light period, this indicated that malate transported from the vacuole was decarboxylated to supply CO₂ to Calvin cycle with high concentration levels. Under the identical conditions, terrestrial C₃ and C₄ species showed no evidence of this type of O₂ evolution (Kawamitsu and Boyer, 1999). The O₂ evolution was decreased with time probably due to depletion of the stored malate. The inset in Fig. 5 indicates the difference in photosynthetic O₂ evolutions between at 3% CO₂ and CO₂ free air. Interestingly, the difference was high at mid-day and low at the beginning and end of light period, suggesting that the activities of Rubisco, a predominant CO₂ fixing enzyme, were not only dependent on carbon dioxide concentration inside the leaf but also regulative by a diurnal rhythm of CAM.

To ensure the effects of very high CO₂ concentration on CO₂ exchange rate in a whole day, we measured diurnal changes in CO₂ exchange rate at 0.0392, 0.1078 and 1.017% CO₂ with a novel gas exchange system (Graan and Boyer, 1990; Kawamitsu and Boyer, 1999) (Fig. 8). Diurnal patterns of CO₂ exchange at 0.0392% CO₂ was a typical CAM mode and Phase IV was clearly identified as in Fig. 5. When CO₂ concentration increased to 0.1078 or 1.017%, CO₂ uptake was increased not only during the light period but also during the dark period (Table 1). Especially at 1.017% CO₂, the CO₂ uptake was significantly enhanced during Phases II

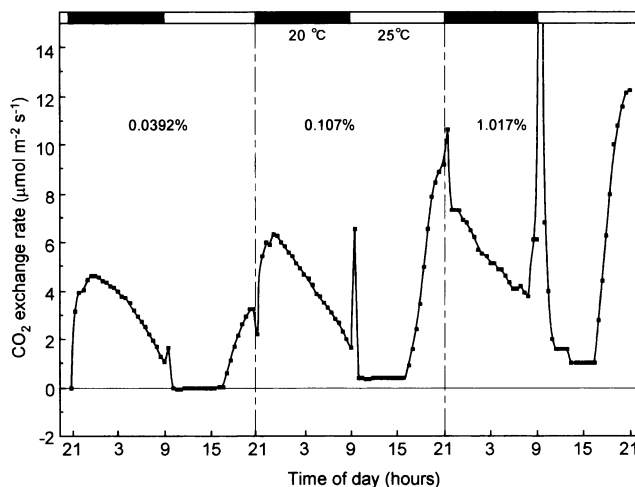


Fig. 8 Effects of CO₂ concentration on the time courses of CO₂ exchange in pineapple. Day/night temperature was set at 25/20°C, and PPFD was 650 μmol m⁻² s⁻¹ during the light period.

Table 1 Effect of CO₂ concentration on CO₂ balance (relative to 0.0392% CO₂) in pineapple.

Phase	CO ₂ concentration (%)		
	0.0392	0.1078	1.017
Total	100	178	272
I	100	133	147
II and III	100	400	2 400
IV	100	400	560

and III because incoming CO₂ from the atmosphere was fixed by Rubisco. Interestingly, CO₂ fixation during the dark period was also increased even though during which PEPCase with higher affinity to CO₂ mainly regulated CO₂ fixation (Kluge and Ting, 1978; Ting, 1994; Winter and Smith, 1996). As in C₄ plants, PEPCase in CAM plants during Phase I was thought to be saturated at near-ambient CO₂ levels (Ting, 1994; Winter and Smith, 1996). However, Zhu et al. (1997) showed that Phase I CO₂ uptake by PEPCase in pineapple was not CO₂ saturated at ambient CO₂ levels, and that the enhancement of CO₂ dark fixation by elevated CO₂ was probably due to the CO₂ un-saturation of PEPCase, the reduced dark respiration and the improvement of the low mesophyll conductance.

In addition to these factors, since the capacity of vacuoles as reservoir in mesophyll cells was substantially restricted, then increments in CO₂ fixation by means of CO₂ enrichment have not been expected to enhance the CO₂ uptake during Phase I (Black, 1986; Ting, 1994). However, as shown in Fig. 8 in our study and some other studies (Zhu et al., 1997, 1999), high CO₂ concentration significantly increased the dark CO₂ fixation. For this, we emphasize again that the CO₂ diffusion will be insufficient at the most inner side of the succulent leaf of pineapple, and only the mesophyll cells near the stomatal cavities actively fix CO₂ from the ambient air. Thus the high CO₂ concentration overcomes the slower CO₂ diffusion in inner mesophyll cells in pineapple.

In CAM plants stomata close with increasing CO₂ (Kawamitsu et al., 1999b), so that the effect of CO₂ enrichment on CO₂ uptake seems to be simultaneously restricted with the stomatal closure. However, even though the stomata are closed at high CO₂, the very high ambient CO₂ enhances CO₂ uptake not only during the light period, but also during the dark period. The increased CO₂ uptake and reduced stomatal conductance under CO₂ enrichment increased the water-use efficiency in pineapple and also other CAM plants (Zhu et al., 1999). Moreover, as mentioned above, the very high CO₂ significantly improves the CO₂ supply to the thicker mesophyll cell, which provides higher capability to fix CO₂.

Based on the photosynthetic O₂ evolution or CO₂ exchange rates presented here, the more increased CO₂ uptake in pineapple at very high CO₂ concentration will be attained both in the light and dark periods. It is necessary, therefore, to carry out the nighttime CO₂ enrichment in pineapple cultivation under greenhouse conditions when windows are closed at night in winter season.

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CAM 植物における明期の光合成の制御
——気相型酸素電極および補償型赤外線 CO₂ 分析装置による評価——

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通常の CO₂ 交換測定装置では評価が不可能な CAM 植物 (パイナップル (*Ananas comosus*) を含む 12 種) の明期における葉内部の光合成活性を, 気相型酸素電極および補償型赤外線 CO₂ 交換測定装置を用いて検討した。5% の CO₂ 濃度下で測定した場合, パイナップルの酸素放出速度は 1 500 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPF D で光飽和し, その最大値は通常のガス交換速度で得られる値の約 10 倍の 60 $\mu\text{mol m}^{-2} \text{s}^{-1}$ であった。また, 飽和光下の酸素放出速度は CO₂ 濃度が 3% までは上昇を続け, 4% 以上では低下した。しかしながら, 他の CAM 植物ではそのような CO₂ 濃度上昇に伴う酸素放出速度の増加は小さかった。次に, 補償型 CO₂ 交換測定装置を用いて, 高 CO₂ 濃度条件下でパイナップルの CO₂ 交換速度を測定したところ, CO₂ の取り込みは明期の Phase III だけではなく暗期の Phase I においても増大することがわかった。以上の結果に基づき, CAM 植物, 特にパイナップルにおける CO₂ 取り込み速度の更なる上昇の可能性を, 気孔の機能やリンゴ酸の貯蔵容量の観点から考察した。